

**STEVE M. BLEVINS, MD**

Assistant Professor of Medicine, Section of General Internal Medicine, Department of Medicine, University of Oklahoma Health Sciences Center, Oklahoma City

RONALD A. GREENFIELD, MD*

Professor of Medicine, Section of Infectious Diseases, Department of Medicine, University of Oklahoma Health Sciences Center and the Oklahoma City Veterans Administration Medical Center

MICHAEL S. BRONZE, MD

Professor of Medicine, Section of Infectious Diseases, Chair of Department of Medicine, University of Oklahoma Health Sciences Center and the Oklahoma City Veterans Administration Medical Center

Blood smear analysis in babesiosis, ehrlichiosis, relapsing fever, malaria, and Chagas disease

ABSTRACT

Blood smear analysis is especially useful for diagnosing five infectious diseases: babesiosis, ehrlichiosis, relapsing fever due to *Borrelia* infection, malaria, and American trypanosomiasis (Chagas disease). It should be performed in patients with persistent or recurring fever or in those who have traveled to the developing world or who have a history of tick exposure, especially if accompanied by hemolytic anemia, thrombocytopenia, or hepatosplenomegaly.

KEY POINTS

In the United States, malaria and American trypanosomiasis principally affect travelers from the developing world.

Babesiosis, ehrlichiosis, and relapsing fever are transmitted by ticks and may produce thrombocytopenia and elevation of liver enzyme levels.

Malaria and babesiosis cause hemolytic anemia and may be associated with hepatomegaly and splenomegaly.

Recurring fever is typical of malaria and *Borrelia* infection.

*Dr. Greenfield has disclosed that he has received honoraria from the Astellas, Bristol-Myers Squibb, Cubicin, Gilead, Ortho-McNeil, and Pfizer corporations for teaching and speaking.

BLOOD SMEAR ANALYSIS, while commonly used to evaluate hematologic conditions, is infrequently used to diagnose infectious diseases. This is because of the rarity of diseases for which blood smear analysis is indicated. Consequently, such testing is often overlooked when it is diagnostically important.

Nonspecific changes may include morphologic changes in leukocytes and erythrocytes (eg, toxic granulations, macrocytosis).¹ And with certain pathogens, identifying organisms in a peripheral blood smear allows for a rapid diagnosis.

This paper discusses the epidemiology, clinical manifestations, laboratory findings, and management of five infectious diseases in which direct visualization of the organism in the blood plays a major diagnostic role. Our intent is to summarize the clinical findings that should prompt blood smear analysis so that these uncommon conditions are not overlooked.

BABESIOSIS

Babesiosis, a tick-borne protozoal disease, occurs principally in the United States and Europe. Of the more than 100 species of *Babesia*, two account for almost all human disease: *B microti* and *B divergens*.

Both species are transmitted by *Ixodes* ticks, although patients often do not recall being bitten. The disease may rarely complicate blood transfusion. Most cases occur from May to September, when tick exposure is

Babesiosis

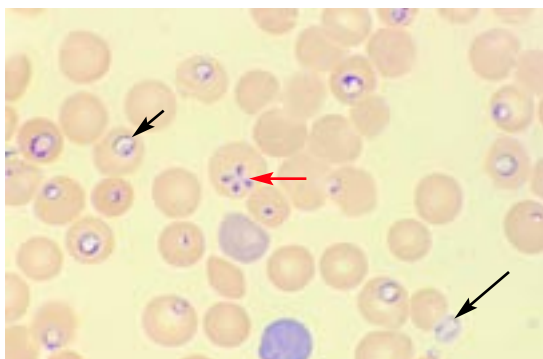


FIGURE 1. *Babesia microti* forming intraerythrocytic rings (short arrow), exoerythrocytic rings (long arrow) and a “Maltese cross” (red arrow) (Giemsa stain, original magnification $\times 100$).

highest. The incubation period varies from 1 to 4 weeks.

Common in the Northeast, usually asymptomatic

B microti infection occurs predominantly in the United States. Rodents, especially the white-footed mouse, are the principal reservoir.^{2,3} Endemic areas, where seropositivity rates range from 4% to 21%, include the coastal areas and islands off of Massachusetts, particularly Cape Cod, Nantucket, and Martha’s Vineyard; the islands near New York City, especially Long Island, Shelter Island, and Fire Island; Block Island, off the coast of Rhode Island; and certain areas in Connecticut.⁴ WA-1, a species that is morphologically identical to *B microti*, is emerging in California and Washington.^{5,6}

Infection with *B microti* is usually asymptomatic. Elderly and immunosuppressed people, especially those without a spleen or with impaired cellular immunity, are more likely to become ill. Symptoms, including fever, malaise, headache, nausea, and generalized aching, may last weeks to months.

About one-fourth of patients with babesiosis are coinfecting with the Lyme disease bacterium (*Borrelia burgdorferi*) and often have more severe illness.³

Hepatomegaly, splenomegaly, jaundice, and dark urine are common findings in patients with symptoms. Severe hemolysis, often accompanied by thrombocytopenia,

leukopenia, and atypical lymphocytosis, is more common in high-risk patients. Hepatic transaminases may be elevated. Urinalysis may show proteinuria and hemoglobinuria. Acute respiratory distress syndrome has been reported in severe cases.^{7–10}

B divergens infection:

A serious but rare disease seen in Europe

B divergens is found mainly in Europe. Altogether, fewer than 50 cases of infection have been reported in France, Spain, Germany, Great Britain, Ireland, Yugoslavia, and the former Soviet Union.^{11,12} Cattle are the principal reservoir of infection.

Infection with *B divergens* causes a rare but devastating disease mainly in asplenic people, usually resulting in coma and death. No cases of subclinical infection have been reported. The clinical course is fulminant, and hemolytic anemia is common.^{2,3}

Suspect babesiosis in endemic areas in cases of prolonged ‘flu’

Babesiosis should be considered when a patient residing in or traveling from an endemic area presents with a prolonged flu-like illness and hemolysis, with or without organomegaly and jaundice.

The diagnosis is made by finding intraerythrocytic parasites on a Giemsa-stained blood smear (FIGURE 1). Thin smears are preferred to thick smears for visualizing the organism. Levels of parasitemia may range from less than 1% to more than 80% of erythrocytes. High levels of parasitemia occur mainly in asplenic patients. At low levels of parasitemia, meticulous blood smear examination is required.

The protozoa may resemble the rings of malaria parasites. Distinguishing traits include exoerythrocytic organisms; the absence of pigmented granules in infected red blood cells; and a “maltese cross,” a rare pattern produced by tetrads of *Babesia* merozoites.¹³ An infected erythrocyte may contain up to eight parasites.

Serologic and polymerase chain reaction tests are useful when the organism is not visible.^{14,15}

Treat patients with severe disease

Most patients with *B microti* infection have a mild illness that resolves without treatment.

Suspect babesiosis in New England in prolonged ‘flu’ and hemolysis

Treatment is recommended for those with severe infection and in those with high-level parasitemia. Agents with consistent activity against *B microti* include clindamycin (Cleocin), azithromycin (Zithromax), atovaquone (Mepron), doxycycline (Vibramycin), and quinine (Quinamm). Combination therapy with either clindamycin and quinine or azithromycin and atovaquone is recommended. *B divergens* infection has been successfully treated with a combination of clindamycin, quinine, and exchange transfusion.¹⁶

■ EHRLICHIOSIS

Ehrlichiosis, nicknamed Rocky Mountain “spotless” fever, is a seasonal, tick-borne disease caused by obligate intracellular bacteria. Bacteria of the genus *Ehrlichia* grow within the cytoplasmic vacuoles of leukocytes and cause mainly zoonotic infections. Several species, especially *Ehrlichia chaffeensis* and *Anaplasma phagocytophilum*, are recognized as human pathogens.^{17,18} *E chaffeensis* infects mononuclear cells, causing a condition known as human monocytic ehrlichiosis (HME). *A phagocytophilum* infects neutrophils, producing a condition called human granulocytic anaplasmosis (HGA).

Deer are the principal reservoir for *E chaffeensis*¹⁹; white-footed mice, other rodents, and deer are the principal reservoirs for *A phagocytophilum*. HME is transmitted by *Dermacentor* and *Ixodes* ticks, and HGA by *Ixodes* ticks. Human infection usually occurs in the spring and summer, when tick exposure is greatest. Co-infection of ticks with the organisms causing Lyme disease or babesiosis may result in simultaneous transmission of these diseases.

More than 1,000 cases of HME have been reported in the southeastern, south-central, and mid-Atlantic regions of the United States.²⁰ The prevalence of HME in the United States appears to follow that of Rocky Mountain spotted fever. Some cases have been described in New England and in the Pacific Northwest. The more than 600 reported cases of HGA have come from Wisconsin, Minnesota, Connecticut, New York, Massachusetts, California, Florida, and Western Europe.^{21,22} The distribution of

HGA follows that of Lyme disease, because the two diseases share the same tick vector.

Acute onset of fever and myalgias

HME and HGA have an incubation period of 1 to 2 weeks. The symptoms are similar and are usually acute, ranging from mild to severe. Most patients have fever, chills, malaise, headache, and myalgias. Many also have nausea, vomiting, cough, and arthralgias. Symptoms are similar to those in Rocky Mountain spotted fever (caused by *Rickettsia rickettsii*), except that rash is uncommon in HME (seen in approximately a third of patients) and rare in HGA.^{23–25} Neurologic findings, such as altered sensorium and neck stiffness, may be accompanied by lymphocytic pleocytosis and elevated protein levels in the cerebrospinal fluid.²⁶ Subclinical and subacute presentations (eg, a fever lasting up to 2 months) are uncommon. No chronic cases have been reported.

The estimated death rate is 1% to 10%, and hospitalization rates are as high as 60%. Most deaths occur in the elderly, often following such complications as congestive heart failure,²⁷ cardiac tamponade, respiratory or renal failure, seizures, and coma. Patients with human immunodeficiency virus infection also have a poor prognosis. Convalescence may be prolonged.

Laboratory abnormalities include leukopenia, thrombocytopenia, and elevated hepatic transaminase levels. Leukopenia may be associated with lymphopenia or neutropenia. Lymphopenia occurs early in the course of illness and is usually followed by an atypical lymphocytosis. Prolonged symptoms are associated with a decreased total neutrophil count and an increased band neutrophil count.²⁸

Suspect ehrlichiosis in endemic areas in patients with fever, leukopenia, or thrombocytopenia

Ehrlichiosis should be suspected when a febrile patient with leukopenia or thrombocytopenia has been exposed to ticks in an endemic area. Even patients whose cell counts and liver enzyme levels are normal should be evaluated if the clinical and epidemiologic situations suggest this disease.

The peripheral blood smear should be

**Ehrlichiosis:
Rocky
Mountain
'spotless' fever**

Ehrlichiosis

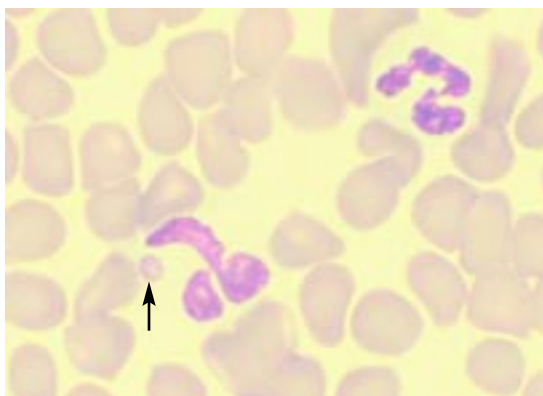


FIGURE 2. Morula of *Anaplasma phagocytophilum* in the cytoplasm of a neutrophil (arrow) (Wright stain, original magnification $\times 100$).

examined for intracytoplasmic inclusions (morulae) within mononuclear cells (for HME) and within neutrophils (for HGA) (FIGURE 2). Blood smear examination is more likely to be positive in patients with HGA than in those with HME. Because the number of infected leukocytes may be low, examination of more than 500 white blood cells on a Wright-stained smear is recommended.²⁹ Buffy coat examination, which allows for concentrated white blood cell analysis, improves the diagnostic yield.

Other diagnostic tests include polymerase chain reaction and serologic assays, which are highly sensitive and specific.^{30,31} Because the organisms are difficult to culture in vitro, blood cultures are not useful diagnostically.

Treatment

Doxycycline 100 mg twice daily for 7 to 10 days is the treatment of choice for both HME and HGA. No role has been defined for fluoroquinolones for treating these diseases. Avoiding ticks and removing ticks promptly are the best preventive strategies.

RELAPSING FEVER

Relapsing fever is an acute febrile illness caused by spirochetes of the genus *Borrelia*. The disease has two forms: tick-borne, in which human infection is zoonotic, and louse-borne, in which humans are the only known reservoir of infection.³²

Few tick-borne cases in the United States

Tick-borne disease is caused by many species of *Borrelia*. Those found in the United States occur in the western mountains and high deserts and plains of the Southwest.³³ Fewer than 30 cases of tick-borne relapsing fever are diagnosed in the United States annually.

Tick-borne relapsing fever is transmitted by soft-bodied argasid ticks (*Ornithodoros* genus), which feed for less than an hour (usually at night) and can survive for years without a blood meal. They stay close to human and animal habitations. Exposure often occurs in cabins, under buildings, in caves, near woodpiles, and in rooms shared with animals. Rodents are the primary animal reservoir. In contrast, most other tick-borne diseases—babesiosis, ehrlichiosis, Lyme disease, Rocky Mountain spotted fever, Colorado tick fever—are transmitted by hard-bodied ixodid ticks, which live in brush and forested areas and attach to passersby, on whom they feed for days if not removed.^{34–36}

Louse-borne disease is endemic in Africa

Louse-borne relapsing fever is caused by a single species, *B recurrentis*, endemic in Ethiopia and Sudan. It may occur sporadically or in epidemics. War, famine, and mass migrations predispose to epidemics with death rates ranging from 30% to 70% if untreated.^{37,38} Disease is spread between humans by the human body louse (*Pediculus humanus*).

Relapsing high fevers

The clinical manifestations of tick-borne and louse-borne relapsing fever are similar, although louse-borne relapsing fever often has a longer incubation period and a longer duration of illness. Bacteremia is heralded by the acute onset of high fever (usually above 39°C [102.2°F]), accompanied by headache, nausea, myalgias, and arthralgias. On average, clinical illness remits in 3 days in tick-borne relapsing fever, but may take 5 to 6 days in louse-borne relapsing fever. Physical findings may include altered sensorium, petechiae, hepatosplenomegaly, and conjunctival suffusion. The fever culminates in a “crisis,” characterized by rigors and a precipitous rise in temperature, pulse, and blood pressure. This is followed by defervescence, diaphoresis, and hypotension. The risk of death

The ticks that carry *Borrelia* can survive for years without a blood meal

is highest during this period and immediately afterwards.

With resolution of the bacteremia, an afebrile period ensues, lasting 4 to 14 days. Fever then recurs, although usually milder, again associated with bacteremia. On average, people with tick-borne relapsing fever have three febrile relapses; those with louse-borne relapsing fever have one.³⁹ Relapse occurs because of antigenic variation, in which a major surface antigen of the spirochete is changed to evade the host's immune system.^{40–42}

***Borrelia* may invade organs and the nervous system**

With each episode of bacteremia, spirochetes may penetrate the brain, heart, liver, eye, or inner ear. Involvement of the central nervous system is more common with tick-borne than with louse-borne relapsing fever. Nervous system involvement may include facial palsy, myelitis, radiculopathy, aphasia, hemiplegia, stupor, or coma.^{43,44} Myocarditis, common in both forms of relapsing fever, portends a poor prognosis.⁴⁵ Invasion of the eye or ear may result in visual impairment or dizziness. Bleeding disorders, manifested by epistaxis, petechiae, and ecchymoses, are typical of louse-borne disease and may be associated with low-grade disseminated intravascular coagulation.⁴⁶ Splenomegaly is more common in louse-borne than in tick-borne disease.

Auxiliary test findings

Laboratory findings include normocytic anemia, leukocytosis, and thrombocytopenia. Liver enzyme levels may be elevated and coagulation tests may be prolonged. Patients with cardiac involvement may have a prolonged QTc interval. Cardiomegaly and pulmonary edema may be seen on chest radiography. The cerebrospinal fluid in patients with neurologic involvement has a mononuclear pleocytosis and a mildly elevated protein concentration.

Suspect if recurrent fever in endemic areas

The diagnosis should be suspected in endemic areas in patients with recurrent fever who have been exposed to ticks or lice. A definitive diagnosis is made by blood smear examination during a febrile period. Spirochetes can

Relapsing fever

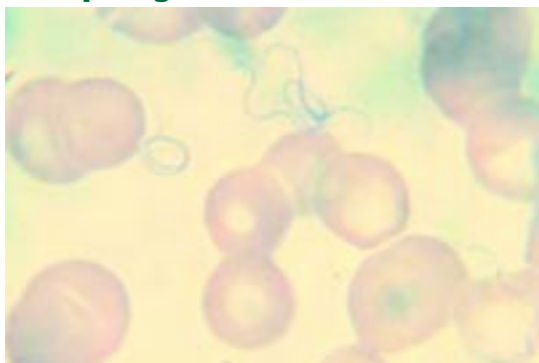


FIGURE 3. Spirochetes of the genus *Borrelia* (Wright stain, original magnification $\times 100$).

be seen on thin or thick smears using Wright and Giemsa stains (FIGURE 3). The organisms are not detectable between febrile episodes. Serologic assays may be unreliable, and false-positive tests for other treponemal illnesses (Lyme disease and syphilis) may occur.

Treatment

Relapsing fever can be successfully treated with tetracycline, penicillin, or erythromycin.⁴⁷ The preferred regimen for a nonpregnant adult with louse-borne relapsing fever is a single 0.5-g dose of tetracycline. In tick-borne relapsing fever, 0.5 g of tetracycline four times daily for 5 to 10 days is recommended. Meningitis or encephalitis is usually treated with parenteral penicillin or ceftriaxone (Rocephin). The death rate in treated disease is usually less than 5%.³⁹ Treatment can induce a Jarisch-Herxheimer reaction (rigors and hypotension, resembling a febrile crisis), and patients must be watched closely for the first 4 hours after the antibiotic is given. Avoiding ticks and practicing good personal hygiene to prevent acquiring lice are the major preventive strategies.

■ MALARIA

Malaria is caused by protozoa of the genus *Plasmodium*. Injected during a mosquito bite, sporozoites enter the bloodstream and travel to the liver. During the asymptomatic hepatic stage, the sporozoites multiply within hepatocytes to form mature schizonts. Within 1 to 2 weeks, the schizonts rupture, releasing thousands of merozoites into the bloodstream. The

With each episode of bacteremia, spirochetes may penetrate the brain, heart, liver, eye, or inner ear

merozoites enter red blood cells, where they mature through the trophozoite stage to become schizonts, which release more merozoites into the blood that may infect other red blood cells. Symptoms occur during this erythrocytic stage, usually 1 to 4 weeks after a mosquito bite.

Four *Plasmodia* species regularly cause human disease.⁴⁸ All can digest hemoglobin and cause hemolysis. *P falciparum* is uniquely dangerous, in part because it can alter the erythrocyte surface and obstruct the microcirculation.^{49,50} Further, *P falciparum* can cause high levels of parasitemia because it can infect red blood cells in all stages of their maturation. In contrast, *P vivax* and *P ovale* invade only reticulocytes, so levels of parasitemia are low (< 1%). *P vivax* and *P ovale* produce dormant liver forms (hypnozoites), which may cause relapses, usually within 3 years of exposure. Falciparum malaria does not have a dormant form, so relapses do not occur. *P malariae* infects only mature red blood cells, producing low levels of parasitemia. A fifth malaria species once thought to be confined to monkeys, *P knowlesi*, is a rare cause of severe human disease.⁵¹

Up to a half-billion cases of malaria occur worldwide each year.⁵² Although the disease predominates among residents of endemic areas, about 30,000 travelers from industrialized countries are infected each year. Fewer than 1,000 cases are reported in the United States annually, which were usually acquired during travel to endemic areas.^{53,54} The risk of transmission varies from region to region, with highest rates (listed in descending order) occurring in Oceania, sub-Saharan Africa, the Indian subcontinent, Southeast Asia, South America, and Central America.⁵⁵ More than 3 million people die of malaria annually, most of them in sub-Saharan Africa.

Malaria is transmitted principally by the bite of the female *Anopheles* mosquito but can also be transmitted by blood transfusion, by the use of contaminated needles, congenitally, and through organ transplantation. Because immigrants to the United States from endemic areas may harbor the parasites for months to years, malaria may rarely be acquired by autochthonous transmission, in which a parasitized individual infects competent vectors, which then bite uninfected persons.⁵⁵ The dis-

ease may also be transmitted from parasitized mosquitoes that arrive on an aircraft ("airport malaria"). There is no animal reservoir for human malaria parasites.

Recurring fever

Fever is universally present, irrespective of the species causing infection. Common symptoms include malaise, chills, headache, myalgias, abdominal pain, night sweats, nausea, and diarrhea. Febrile episodes, initially frequent and irregular, may become regular, producing temperature spikes every second or third day depending on the species. The severity of the paroxysms usually diminishes over time. Eventually, anemia, thrombocytopenia, jaundice, splenomegaly, and hepatomegaly occur.

Falciparum malaria may result in pulmonary edema, renal failure, gastroenteritis, bleeding, or hypoglycemia. Cerebral disease, presenting as altered sensorium or seizures, can be fatal.⁵⁶ Death, common in untreated patients, correlates with the degree of parasitemia.⁵⁷ Severe disease often occurs in children, pregnant women, asplenic patients, and nonimmune adults.

Infection with *P vivax* or *P ovale* does not produce the microvascular complications of falciparum malaria, and thrombocytopenia is less common. *P malariae* infection may lead to immune complex deposition and nephrotic syndrome. Symptoms, although mild, are often prolonged.

Consider malaria in travelers with recurring fever

Malaria should be suspected in febrile patients who have traveled to an endemic region. Even in nontravelers, malaria should be considered in the differential diagnosis of fever of unknown origin. The diagnosis is made by detecting intraerythrocytic organisms on a Giemsa-stained blood smear (FIGURE 4).^{58,59} Thick smears are especially sensitive, but thin smears are useful for identifying the species and for estimating the level of parasitemia. The first smear is positive in more than 90% of cases. If negative, additional smears should be obtained every 6 to 12 hours for 48 hours because of the cyclical nature of the parasitemia. Two hundred to 300 oil-immersion fields should be viewed before a smear is considered negative.⁶⁰

Up to a half-billion cases of malaria occur worldwide each year

Antigen-capture test kits are useful for rapid diagnosis. These portable devices detect parasitic antigens from a drop of blood in just 15 minutes.⁶¹ Polymerase chain reaction techniques offer high sensitivity and specificity but are more expensive and less available.⁶²

The management of patients with suspected malaria should be done by experienced health care providers. Recommendations about malaria treatment are beyond the scope of this review. The choice of antimalarial drugs requires knowledge of the regional distribution of drug resistance and the adverse effects of the agents. Patients with non-falciparum malaria rarely require hospitalization.

Malaria is prevented by mosquito avoidance and chemoprophylaxis.

■ AMERICAN TRYPANOSOMIASIS (CHAGAS DISEASE)

American trypanosomiasis is a zoonotic, protozoal disease caused by *Trypanosoma cruzi*.

The parasite is transmitted by bloodsucking triatomine insects, or “kissing bugs.”⁶³ These insects favor mud-brick and clay houses, where they live in wall cracks, under furniture, and behind pictures. The insects acquire the organism by feeding on infected animals or on humans that have circulating trypomastigotes. The organisms then multiply in the gut of the insects and are transmitted to a second vertebrate host as the insect defecates following a blood meal. The parasite then enters the body through the skin, conjunctivae, or mucous membranes. After entering the body, the parasite disseminates through the bloodstream, invading many cell types, especially muscle and nerve.

American trypanosomiasis occurs in Central and South America, Mexico, and the southern United States. Between 16 and 18 million people are infected with the parasite, and nearly 50,000 die annually, usually from cardiac complications. Once confined to rural areas, the disease is now common in cities. The majority of reported cases come from Brazil. Only a few cases have been reported in the United States, but an estimated 50,000 to 100,000 immigrants are thought to be infected.^{64–68}

Transmission of the parasite may also

Malaria

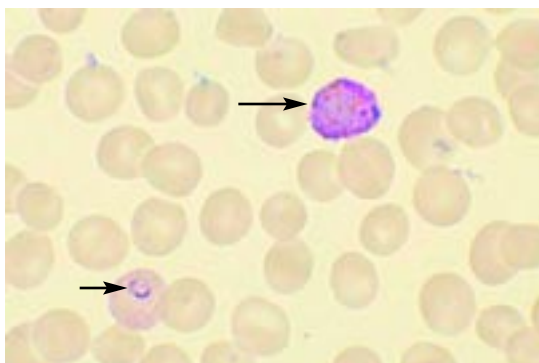


FIGURE 4. *Plasmodium vivax* organisms: intraerythrocytic ring (short arrow); mature trophozoite (long arrow) (Giemsa stain, original magnification $\times 100$).

occur with blood transfusion, with organ transplantation, and congenitally.^{63,69,70} An increase in transfusion-related cases is expected in the United States because of an influx of migrant workers from Mexico and Central America.

Acute phase ranges from asymptomatic to multiorgan involvement

Most infected people are asymptomatic carriers. Only 1% become acutely ill, but up to a third of those infected may develop chronic symptoms decades later. Acute Chagas disease, usually seen in children, is characterized by fever, malaise, and anorexia, often accompanied by vomiting, diarrhea, and rash.^{64,71,72} The heart, liver, spleen, and lymph nodes become enlarged. A red and indurated nodule or furuncle (chagoma) appears at the inoculation site. If inoculation occurs across the conjunctivae, painless edema of the palpebrae and periocular tissues may be observed (Romaña sign). Generalized lymphadenopathy and hepatosplenomegaly may also be seen.

Myocarditis and meningoencephalitis occur in some cases of acute Chagas disease. Myocardial inflammation may extend to the pericardium, causing pericardial effusion, and to the endocardium, causing thrombus formation. All cardiac chambers become enlarged and the conduction system is disrupted.⁷³ Brain damage from meningoencephalitis usually occurs in infants and young children, and may result in death.

Even in nontravelers, malaria should be considered for fever of unknown origin

Chagas disease

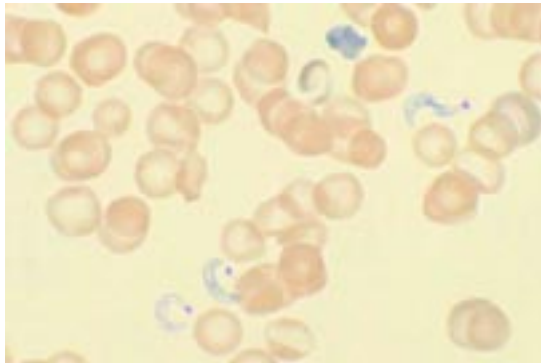


FIGURE 5. C-shaped trypomastigotes of *Trypanosoma cruzi* (Wright stain, original magnification $\times 100$).

This acute phase lasts 4 to 8 weeks and is characterized by profound parasitemia, tissue invasion, and inflammation. Following the acute phase, an asymptomatic latent or indeterminate phase lasting 10 to 40 years occurs.⁷⁴ Less than half of those in clinical latency enter a chronic phase of disease.

Severe chronic phase may occur decades later

The chronic phase, which occurs years to decades after the initial infection, is characterized by cardiac, esophageal, and colonic enlargement. Cardiac involvement is associated with congestive heart failure, arrhythmias, and cardiac arrest. Intracardiac thrombi may embolize, causing systemic and pulmonary infarctions.^{71–73,75} Enlargement of the esophagus is associated with dysphagia, chest pain, weight loss, and sometimes perforation or aspiration-related pneumonitis. Colonic enlargement may result in constipation, abdominal

distention, and intestinal obstruction. Sometimes, the small bowel, ureters, and bronchi become dilated as well. These findings are the result of low-grade parasitemia, tissue inflammation, and immune-mediated disruption of the microvasculature.^{71,76}

Diagnosis: Protozoa evident in acute phase

In the acute phase, Chagas disease is readily diagnosed by examining a fresh drop of unstained blood. The rapid protozoal movements can be seen at $400\times$ magnification. At least 100 fields should be viewed. Stained blood smears are less sensitive, but may allow visualization of the nucleus, the kinetoplast, and flagellum (FIGURE 5). Blood concentration techniques may be used when the initial examination is unrevealing.

Serologic tests are most useful for diagnosing chronic Chagas disease.⁷⁷ The organism can be identified by polymerase chain reaction, but the sensitivity of this test is highly variable.

Treatment reduces symptoms

Two drugs are used to treat American trypanosomiasis. Nifurtimox (Lampit) reduces symptom duration and severity, as well as mortality rates, in acute and congenital Chagas disease; however, fewer than 75% of patients have a parasitologic cure, and adverse effects limit tolerability.⁷⁸ Benznidazole (Rochagan, Radanil) has an efficacy similar to that of nifurtimox and is considered the drug of choice in Latin America. These drugs are not commercially available in the United States. Therapy may help during the indeterminate phase but is rarely effective for chronic Chagas disease. Treatment of Chagas disease with triazoles is under evaluation. ■

The parasite of American trypanosomiasis (Chagas disease) is transmitted by triatomine insects ('kissing bugs')

REFERENCES

1. Kroft SH. Infectious diseases manifested in the peripheral blood. Clin Lab Med 2002; 22:253–277.
2. Meldrum SC, Birkhead GS, White DJ, Benach JL, Morse DL. Human babesiosis in New York State: an epidemiological description of 136 cases. Clin Infect Dis 1992; 15:1019–1023.
3. Pruthi RK, Marshall WF, Wiltse JC, Persing DH. Human babesiosis. Mayo Clin Proc 1995; 70:853–862.
4. Eskow ES, Krause PJ, Spielman A, Freeman K, Aslanzadeh J. Southern extension of the range of human babesiosis in the eastern United States. J Clin Microbiol 1999; 37:2051–2052.
5. Quick RE, Herwaldt BL, Thomford JW, et al. Babesiosis in Washington State: a new species of *Babesia*? Ann Intern Med 1993; 119:284–290.
6. Persing DH, Herwaldt BL, Glaser C, et al. Infection with a babesia-like organism in northern California. N Engl J Med 1995; 332:298–303.
7. Gorenflot A, Moubri K, Precigout E, Carcy B, Schetters TP. Human babesiosis. Ann Trop Med Parasitol 1998; 92:489–501.
8. Villar BF, White DJ, Benach JL. Human babesiosis. Prog Clin Parasitol 1991; 2:129–143.
9. Sun T, Tenenbaum MJ, Greenspan J, et al. Morphologic and clinical observations in human infection with *Babesia microti*. J Infect Dis 1983; 148:239–248.
10. Benach JL, Habicht GS. Clinical characteristics of human babesiosis. J Infect Dis 1981; 144:481.
11. Boustani MR, Gelfand JA. Babesiosis. Clin Infect Dis 1996;

- 22:611–615.
12. Meer-Scherrer L, Adelson M, Mordechai E, Lottaz B, Tilton R. *Babesia microti* infection in Europe. *Curr Microbiol* 2004; 48:435–437.
13. Krause PJ. Babesiosis diagnosis and treatment. *Vector Borne Zoonotic Dis* 2003; 3:45–51.
14. Persing DH, Mathiesen D, Marshall WF, et al. Detection of *Babesia microti* by polymerase chain reaction. *J Clin Microbiol* 1992; 30:2097–2103.
15. Chisholm ES, Sulzer AJ, Ruebush TK 2nd. Indirect immunofluorescence test for human *Babesia microti* infection: antigen specificity. *Am J Trop Med Hyg* 1986; 35:921–925.
16. Berry A, Morassin B, Kamar N, Magnaval JF. Clinical picture: human babesiosis. *Lancet* 2001; 357:341.
17. Maeda K, Markowitz N, Hawley R, Ristic M, Cox D, McDade JE. Human infection with *Ehrlichia canis*, a leukocytic rickettsia. *N Engl J Med* 1987; 316:853–856.
18. Chen SM, Dumler JS, Bakken JS, Walker DH. Identification of a granulocytotropic *Ehrlichia* species as the etiologic agent of human disease. *J Clin Microbiol* 1994; 32:589–595.
19. Lockhart JM, Davidson WR, Stallknecht DE, Dawson JE, Howerth EW. Isolation of *Ehrlichia chaffeensis* from wild white-tailed deer (*Odocoileus virginianus*) confirms their role as natural reservoir hosts. *J Clin Microbiol* 1997; 35:1681–1686.
20. Stone JH, Dierberg K, Aram G, Dumler JS. Human monocytic ehrlichiosis. *JAMA* 2004; 292:2263–2270.
21. Bakken JS, Dumler JS. Human granulocytic ehrlichiosis. *Clin Infect Dis* 2000; 31:554–560.
22. Dumler JS, Bakken JS. Ehrlichial diseases of humans: emerging tick-borne infections. *Clin Infect Dis* 1995; 20:1102–1110.
23. Everett ED, Evans KA, Henry RB, McDonald G. Human ehrlichiosis in adults after tick exposure. Diagnosis using polymerase chain reaction. *Ann Intern Med* 1994; 120:730–735.
24. Fishbein DB, Dawson JE, Robinson LE. Human ehrlichiosis in the United States, 1985 to 1990. *Ann Intern Med* 1994; 120:736–743.
25. Bakken JS, Krueth J, Wilson-Nordskog C, Tilden RL, Asanovich K, Dumler JS. Clinical and laboratory characteristics of human granulocytic ehrlichiosis. *JAMA* 1996; 275:199–205.
26. Ratnasamy N, Everett ED, Roland WE, McDonald G, Caldwell CW. Central nervous system manifestations of human ehrlichiosis. *Clin Infect Dis* 1996; 23:314–319.
27. Vanek NN, Kazi S, Cepero NM, Tang S, Rex JH. Human ehrlichiosis causing left ventricular dilation and dysfunction. *Clin Infect Dis* 1996; 22:386–387.
28. Bakken JS, Aguero-Rosenfeld ME, Tilden RL, et al. Serial measurements of hematologic counts during the active phase of human granulocytic ehrlichiosis. *Clin Infect Dis* 2001; 32:862–870.
29. Hamilton KS, Standaert SM, Kinney MC. Characteristic peripheral blood findings in human ehrlichiosis. *Mod Pathol* 2004; 17:512–517.
30. Dawson JE, Fishbein DB, Eng TR, Redus MA, Green NR. Diagnosis of human ehrlichiosis with the indirect fluorescent antibody test: kinetics and specificity. *J Infect Dis* 1990; 162:91–95.
31. Walls JJ, Caturegli P, Bakken JS, Asanovich KM, Dumler JS. Improved sensitivity of PCR for diagnosis of human granulocytic ehrlichiosis using epank1 genes of *Ehrlichia phagocytophila*-group ehrlichiae. *J Clin Microbiol* 2000; 38:354–356.
32. Barbour AG. Relapsing fever. In: Goodman JL, Dennis DT, Sonenshine DE, editors. *Tick-Borne Diseases of Humans*. Washington, DC: ASM Press; 2005:268.
33. Barbour AG, Hayes SF. Biology of *Borrelia* species. *Microbiol Rev* 1986; 50:381–400.
34. Hoogstraal H. Ticks and spirochetes. *Acta Trop* 1979; 36:133–136.
35. Sonenshine D. *Biology of Ticks*. New York, NY: Oxford University Press; 1991.
36. Felsenfeld O. *Borrelia*; Strains, Vectors, Human, and Animal Borreliosis. St Louis, MO: Warren H Greene, Inc; 1971.
37. Bryceson AD, Parry EH, Perine PL, Warrell DA, Vukotich D, Leithhead CS. Louse-borne relapsing fever. *Q J Med* 1970; 39:129–170.
38. Judge DM, Samuel I, Perine PL, Vukotic D, Ababa A. Louse-borne relapsing fever in man. *Arch Pathol* 1974; 97:136–170.
39. Southern P, Sanford J. Relapsing fever. A clinical and microbiological review. *Medicine* 1969; 48:129–149.
40. Barbour AG. Antigenic variation of a relapsing fever *Borrelia* species. *Annu Rev Microbiol* 1990; 44:155–171.
41. Stoenner HG, Dodd T, Larsen C. Antigenic variation of *Borrelia hermsii*. *J Exp Med* 1982; 156:1297–1311.
42. Barbour AG. Immunobiology of relapsing fever. *Contrib Microbiol Immunol* 1987; 8:125–137.
43. Scott R. Neurological complications of relapsing fever. *Lancet* 1944; 247:436–438.
44. Cadavid D, Barbour AG. Neuroborreliosis during relapsing fever: review of the clinical manifestations, pathology, and treatment of infections in humans and experimental animals. *Clin Infect Dis* 1998; 26:151–164.
45. Wengrower D, Knobler H, Gillis S, Chajek-Shaul T. Myocarditis in tick-borne relapsing fever. *J Infect Dis* 1984; 149:1033.
46. Perine PL, Parry EH, Vukotich D, Warrell DA, Bryceson AD. Bleeding in louse-borne relapsing fever. I. Clinical studies in 37 patients. *Trans R Soc Trop Med Hyg* 1971; 65:776–781.
47. Rhee KY, Johnson WD. *Borrelia* species (relapsing fever). In: Mandell GL, Bennett JE, Dolin R, editors. *Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases*. 6th ed. Philadelphia, PA: Elsevier Churchill Livingstone; 2005:2795–2798.
48. Greenwood BM, Bojang K, Whitty CJ, Targett GA. Malaria. *Lancet* 2005; 365:1487–1498.
49. Newbold C, Craig A, Kyes S, Rowe A, Fernandez-Reyes D, Fagan T. Cytoadherence, pathogenesis and the infected red cell surface in *Plasmodium falciparum*. *Int J Parasitol* 1999; 29:927–937.
50. Oh SS, Chishti AH, Palek J, Liu SC. Erythrocyte membrane alterations in *Plasmodium falciparum* malaria sequestration. *Curr Opin Hematol* 1997; 4:148–154.
51. Singh B, Kim Sung L, Matusop A, et al. A large focus of naturally acquired *Plasmodium knowlesi* infections in human beings. *Lancet* 2004; 363:1017–1024.
52. Fairhurst RM, Wellems TE. *Plasmodium* species (malaria). In: Mandell GL, Bennett JE, Dolin R, editors. *Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases*. 6th ed. Philadelphia, PA: Elsevier Churchill Livingstone; 2005:3121,3122.
53. Filler S, Causer LM, Newman RD, et al; Centers for Disease Control and Prevention (CDC). Malaria surveillance—United States, 2001. *MMWR Surveill Summ* 2003; 52:1–14.
54. Olliaro P, Cattani J, Wirth D. Malaria, the submerged disease. *JAMA* 1996; 275:230–233.
55. Kain KC, Keystone JS. Malaria in travelers. Epidemiology, disease, and prevention. *Infect Dis Clin North Am* 1998; 12:267–284.
56. World Health Organization, Division of Control of Tropical Diseases. Severe and complicated malaria. *Trans R Soc Trop Med Hyg* 1990; 84(suppl 2):1–65.
57. Field JW. Blood examination and prognosis in acute falciparum malaria. *Trans R Soc Trop Med Hyg* 1949; 43:33–48.
58. Microscopic procedures for diagnosing malaria. Appendix. *MMWR Surveill Summ* 2003; 52:15–16.
59. Lema OE, Carter JY, Nagelkerke N, et al. Comparison of five methods of malaria detection in the outpatient setting. *Am J Trop Med Hyg* 1999; 60:177–182.
60. White NJ. The treatment of malaria. *N Engl J Med* 1996; 335:800–806.
61. Moody A. Rapid diagnostic tests for malaria parasites. *Clin Microbiol Rev* 2002; 15:66–78.
62. Makler MT, Palmer CJ, Ager AL. A review of practical techniques for the diagnosis of malaria. *Ann Trop Med Parasitol* 1998; 92:419–433.

63. **Kirchhoff LV.** *Trypanosoma* species (American trypanosomiasis, Chagas' disease): Biology of trypanosomes. In: Mandell GL, Bennett JE, Dolin R, editors. *Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases*. 6th ed. Philadelphia, PA: Elsevier Churchill Livingstone; 2005:3157–3158.
64. **Barrett MP, Burchmore RJ, Stich A, et al.** The trypanosomiasis. *Lancet* 2003; 362:1469–1480.
65. Chagas disease. Interruption of transmission. *Wkly Epidemiol Rec* 1997; 71:1
66. Control of Chagas disease. Report of a WHO Expert Committee. *World Health Organ Tech Rep Ser* 1991; 811:1–95.
67. **Kirchhoff LV.** American trypanosomiasis (Chagas' disease). In: Guerrant R, Walker DH, Weller PF, editors. *Tropical Infectious Diseases: Principles, Pathogens and Practice*; vol. 1. Philadelphia, PA: Churchill Livingstone; 1999:785.
68. **Kirchhoff LV.** American trypanosomiasis (Chagas' disease). *Gastroenterol Clin North Am* 1996; 25:517–533.
69. **Wanderley DM, Corrêa FM.** Epidemiology of Chagas' heart disease. *Sao Paul Med J* 1995; 113:742–749.
70. **Schmunis GA.** *Trypanosoma cruzi*, the etiologic agent of Chagas' disease: status in the blood supply in endemic and nonendemic countries. *Transfusion* 1991; 31:547–557.
71. **Köberle F.** Chagas' disease and Chagas' syndromes: the pathology of American trypanosomiasis. *Adv Parasitol* 1968; 6:63–116.
72. **Dias E, Laranja FS, Miranda A, Nobrega G.** Chagas' disease; a clinical, epidemiologic, and pathologic study. *Circulation* 1956; 14:1035–1060.
73. **Prata A, Andrade Z, Guimaraes AC.** Chagas' heart disease. In: Shaper AG, Hutt MS, Fejfar Z, editors. *Cardiovascular disease in the tropics*. London, England: British Medical Association; 1974:264.
74. **Dias JC.** The indeterminate form of human chronic Chagas' disease. A clinical epidemiological study. *Rev Soc Bras Med Trop* 1989; 22:147–156.
75. **Lopes ER, Tafuri WL.** Involvement of the autonomic nervous system in Chagas' heart disease. *Rev Soc Bras Med Trop* 1983; 16:206.
76. **Meneghelli UG.** Chagas' disease: a model of denervation in the study of digestive tract motility. *Braz J Med Biol Res* 1985; 18:255–264.
77. **Pirard M, Iihoshi N, Boelaert M, Basanta P, Lopez F, Van der Stuyft P.** The validity of serologic tests for *Trypanosoma cruzi* and the effectiveness of transfusional screening strategies in a hyperendemic region. *Transfusion* 2005; 45:554–561.
78. **Kirchhoff LV.** *Trypanosoma* species (American trypanosomiasis, Chagas' disease): Biology of trypanosomes. In: Mandell GL, Bennett JE, Dolin R, editors. *Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases*. 6th ed. Philadelphia, PA: Elsevier Churchill Livingstone; 2005:3162–3163.

ADDRESS: Steve M. Blevins, MD, Department of Medicine, University of Oklahoma Health Sciences Center, WP 1160, 920 Stanton Young Boulevard, PO Box 26901, Oklahoma City, OK 73190; e-mail: Steve-Blevins@ouhsc.edu.