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Clostridium difficile-associated disease: New challenges from an established pathogen

■ ABSTRACT

Clostridium difficile-associated disease (CDAD) can range from uncomplicated diarrhea to sepsis and even death. CDAD rates and severity are increasing, possibly due to a new strain. Transmission of *C difficile* occurs primarily in health care facilities via the fecal-oral route following transient contamination of the hands of health care workers and patients; contamination of the patient care environment also plays an important role.

■ KEY POINTS

A recently identified strain of *C difficile* that has caused numerous outbreaks of clinically severe disease in North America and Europe produces 16 times more toxin A and 23 times more toxin B than other strains.

Since nosocomial CDAD is almost always associated with antimicrobial use, one should avoid unnecessary and inappropriate antimicrobial therapy.

If a patient has CDAD, the clinician must vigilantly monitor for disease progression and follow infection control guidelines to prevent spread to other patients.

Important principles in treating CDAD include stopping the offending antimicrobial agent if possible, giving metronidazole or vancomycin orally for no less than 10 days, and following patients closely for any signs of clinical progression during therapy.

C *LOSTRIDIUM DIFFICILE*-ASSOCIATED DISEASE (CDAD) is increasing in incidence and severity and may be becoming more difficult to treat. Recent reports of a more virulent and possibly more resistant strain of *C difficile*'s causing epidemics in both the United States and Canada have heightened clinicians' awareness of CDAD, emphasizing the importance of early recognition and appropriate treatment.

In this article, we review the current state of knowledge concerning the epidemiology, pathogenesis, clinical presentation, diagnosis, treatment, and prevention of CDAD.

■ CASE REPORT

A 37-year-old man presented to the emergency department because of diffuse abdominal pain and nonbloody diarrhea. One day earlier he had been discharged from the hospital, where he had received ceftriaxone and azithromycin for 7 days for bronchitis. Within hours after going home he passed numerous liquid brown stools; by evening he had become disoriented and an ambulance was called. When he arrived, emergency personnel gave him naloxone for a possible drug overdose, although his fiancé reported that he had taken only one dose each of hydromorphone and lorazepam since returning home (later confirmed by pill count).

His medical history included chronic obstructive pulmonary disease, depression, chronic back pain, and tobacco use. Medications included a fentanyl patch 75 µg/hour every 3 days, gabapentin 600 mg three times a day, hydromorphone 4 mg every 4 hours as needed,

C difficile-associated disease: Resected colonic mucosa

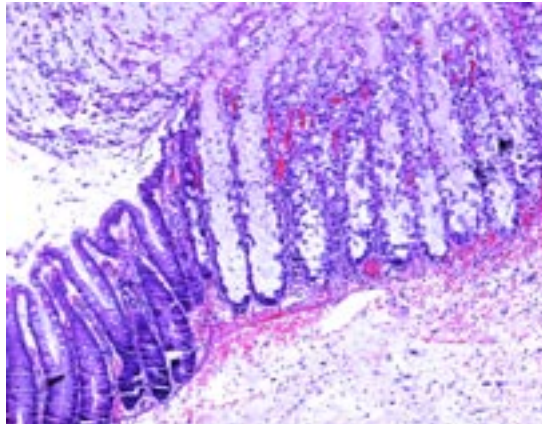


FIGURE 1. Photomicrograph of a hematoxylin and eosin stain of the case patient's colonic mucosa just on the edge of the pseudomembranous lesions. Intact mucosa appears to the left with normal architecture of deep crypts and villi. On the right, destruction of the mucosa with severe inflammatory response extending deep into the lamina propria and expulsion of mucous and cellular debris from the crypts into the lumen of the large intestine, giving the appearance of a volcanic eruption. It is this expelled material that forms the pseudomembranes.

PHOTOMICROGRAPH COURTESY OF JEANNETTE GUARNER, MD,
US CENTERS FOR DISEASE CONTROL AND PREVENTION.

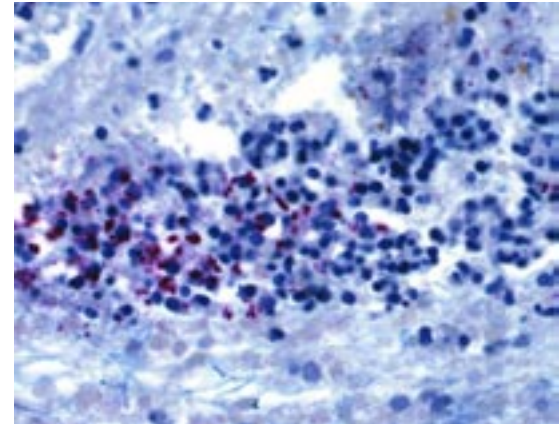


FIGURE 2. Photomicrograph of immunohistochemical stain of the pseudomembrane. This stain uses genus-specific antibodies and demonstrates numerous *Clostridium* species within the pseudomembrane.

PHOTOMICROGRAPH COURTESY OF JEANNETTE GUARNER, MD,
US CENTERS FOR DISEASE CONTROL AND PREVENTION.

**Pseudomem-
branous colitis
is considered
pathognomonic
for C difficile**

lorazepam 1 mg three times a day, and prednisone in tapering doses. He had no known drug allergies and did not use alcohol or illicit drugs.

Laboratory values: white blood cell count $100 \times 10^9/L$, hematocrit 62.3%, sodium 125 mmol/L, potassium 6.6 mmol/L, CO_2 13 mmol/L, and metabolic acidosis.

An abdominal radiographic series showed no evidence of obstruction.

The patient was admitted to the intensive care unit and received fluids and pharmacologic support for hypotension, and metronidazole (Flagyl) 500 mg intravenously. Results of computed tomography of the abdomen were consistent with toxic megacolon.

The patient underwent emergent exploratory laparotomy, which revealed a swollen, edematous colon with pseudomembranes; a subtotal colectomy and ileostomy were performed. After surgery, he was given a second dose of intravenous metronidazole plus

intravenous ciprofloxacin and vancomycin per rectum. Three days after surgery, the patient developed ventricular fibrillation that did not respond to several resuscitation attempts, and he died.

Discussion

Although no testing for *C difficile* was performed before the patient died, histopathologic findings in the resected colon and in the patient's rectum on autopsy were consistent with pseudomembranous colitis, a condition considered pathognomonic for *C difficile* (FIGURE 1). Moreover, an immunohistochemical stain for *Clostridium* species demonstrated numerous organisms within the pseudomembranes (FIGURE 2).

Factors contributing to CDAD and death in this patient include his receiving antimicrobial agents and proton-pump inhibitors, both of which are risk factors for CDAD.¹⁻⁴ Additionally, he received narcotics, which may be a risk factor for toxic megacolon owing to their antiperistaltic effects.^{5,6}

This tragic death of a young, otherwise healthy man illustrates the serious potential complications of CDAD and the importance of preventing and controlling it.

■ INCREASING IN INCIDENCE AND SEVERITY

C difficile is a gram-positive, spore-forming anaerobic bacillus that was first linked to disease in 1978, when it was identified as the causative agent of pseudomembranous colitis.^{7,8} It has been associated with gastrointestinal infections ranging in severity from asymptomatic colonization to severe diarrhea, pseudomembranous colitis, toxic megacolon, intestinal perforation, and death.^{9–11}

C difficile toxins can be found in the stool of 15% to 25% of patients with antibiotic-associated diarrhea and more than 95% of patients with pseudomembranous colitis.¹²

In US hospitals participating in the National Nosocomial Infections Surveillance System,¹³ there were an average of 12.2 reported cases of CDAD per 10,000 patient-days in the years 1987 to 1998. Rates were significantly higher in teaching than in non-teaching hospitals (13.0 vs 11.7 cases per 10,000 patient-days), in medical than in surgical services, and in winter months than in nonwinter months.

Data from the US Centers for Disease Control and Prevention (CDC) reveal that hospitalizations with a discharge diagnosis of CDAD have significantly increased from 31 per 100,000 population in 1996 to 61 per 100,000 in 2003.¹⁴

Of patients who contracted CDAD in hospitals or nursing homes, 0.6% to 1.5% died, and CDAD was either the direct or indirect cause of death.^{15,16} CDAD has been estimated to cost an additional \$3,669 to \$7,234 per patient hospitalization.^{17,18} Moreover, the severity of observed disease may also be increasing, with an attributable 1-year mortality rate approaching 17% in one study.¹⁹

Risk factors for CDAD

Antibiotic therapy. More than 90% of health-care-associated *C difficile* infections occur after or during antimicrobial therapy.^{1,20}

Almost all antimicrobial agents except for aminoglycosides have been associated with CDAD. A meta-analysis by Bignardi¹ suggests that broad-spectrum antimicrobial agents, which have a greater effect on the normal intestinal flora, are more likely to lead to CDAD. However, several later studies found

fluoroquinolones to be more strongly linked to CDAD than any other antimicrobial agents, including clindamycin and beta-lactam/beta-lactamase inhibitors.^{3,21,22} The risk is also greater when patients receive multiple antimicrobial agents and undergo a longer course of therapy.¹

Other risk factors (cited in at least three studies) are:

- Age greater than 65 years
- Severe underlying illness
- Nasogastric intubation
- Antiulcer medications. (There is conflicting evidence regarding the role of proton-pump inhibitors and histamine receptor antagonists in CDAD.^{2–4})
- Longer hospital stay.¹

Specific populations appear to be at greater risk for developing CDAD than the general population. Most cases of CDAD occur in health care settings,²³ as do most CDAD outbreaks.^{24,25} Among hospitalized patients, several studies have found that medical patients are at significantly higher risk of CDAD than surgical patients.^{2,13}

Additionally, *C difficile* is the most common infectious cause of acute diarrheal illness in long-term-care facilities.^{26,27} Even when an outbreak is not going on, the prevalence of *C difficile* colonization in long-term-care facilities ranges from 4% to 20%,²⁶ compared with less than 3% in healthy adults.^{12,20} Compared with the general population, long-term care residents also are older and receive more antibiotics and antacids—all of which are known risk factors for CDAD. These additional risk factors make it difficult to determine which factors contribute most to the increased risk.

Neonates also have more *C difficile* colonization, with cited rates ranging from 5% to 70%.²⁰ Paradoxically, neonates carrying toxigenic strains of *C difficile* are much less likely than adults to develop symptomatic disease. The reason, based on observations in rabbits, may be that neonates lack receptors for toxin A in their immature enterocytes.²⁸

■ HOW *C DIFFICILE* CAUSES DISEASE

For *C difficile* to establish itself and proliferate in the colonic mucosa, the normal flora of the

Almost all antibiotics except aminoglycosides have been associated with CDAD

colon must be disrupted (as with antimicrobials) and *C difficile* must be ingested (FIGURE 3). Although these events need not necessarily occur in that order,²⁰ once both of them occur, the patient can become colonized or develop CDAD.

Toxins are essential for disease

It is unclear why some patients develop disease and others do not; however, toxin production is essential for disease to occur.

C difficile's primary virulence factors are toxins A and B, which are responsible for inflammation, fluid and mucous secretion, and mucosal damage (FIGURE 1), which lead to diarrhea or colitis.²³

A recently identified strain of *C difficile*, designated North American pulsed-field gel electrophoresis type 1 (NAP 1), has caused numerous outbreaks of clinically severe disease in North America and Europe. NAP 1 produces 16 times more toxin A and 23 times more toxin B than other strains,^{29,30} possibly due to a deletion in a negative regulatory gene.³⁰ In addition, NAP 1 produces a third toxin, known as binary toxin, although its significance is unknown. This new strain is resistant to both gatifloxacin and moxifloxacin, which is a new finding compared with historical strains.

Colonization, immunity

Only toxigenic strains of *C difficile* produce clinical disease, but toxin production does not guarantee symptomatic progression.²³ Other host factors can influence the clinical presentation, such as preexisting colonization with *C difficile* and humoral immunity.

Some suggest that colonization with *C difficile* can actually protect against symptomatic disease,³¹ due to the development of immunity. Kyne and colleagues³² demonstrated that asymptomatic carriers had significantly greater antibody responses to toxin A than those who developed nosocomial CDAD.

CLINICAL PRESENTATION VARIES

The incubation period from ingestion of *C difficile* to manifestation of disease has not been established. Symptoms can appear immediately after beginning antimicrobial therapy, or

they may not develop until several weeks after it is completed.²³ In one study of cancer outpatients,³³ the median interval from hospital discharge to CDAD diagnosis was 20.3 days (range 2–60 days)—a considerable delay in disease onset.

The clinical presentation of *C difficile* is a continuum that includes asymptomatic carriage, diarrhea, colitis, pseudomembranous colitis, and fulminant colitis.²³

Mild disease

Most often, CDAD presents as mild to moderate nonbloody diarrhea, sometimes accompanied by low abdominal cramping. Systemic symptoms are typically absent, and physical examination is remarkable only for mild abdominal tenderness.

Severe disease

Colitis, in contrast, tends to present with more severe symptoms, including profuse watery diarrhea and abdominal pain and distention. Fever, nausea, and dehydration are often present. There may be occult blood in the stool, but hematochezia is rare. Sigmoidoscopy reveals a characteristic membrane with adherent yellow plaques, usually in the distal colon, although occasionally it can be confined to the proximal colon and can be missed on examination.

Once severe or systemic symptoms develop, appropriate treatment is crucial to prevent progression to more severe disease.

Patients with severe colitis are at increased risk of developing paralytic ileus and toxic megacolon.²³ These may lead to a paradoxical decrease in diarrhea. Such severe cases may also present as fulminant colitis, with an acute abdomen and systemic symptoms such as fever and tachycardia, as in our case presentation. Such complications require an immediate surgical consult. Of 11 patients with toxic megacolon, 7 (64%) needed surgery, and once patients undergo surgery for complications of CDAD, the mortality rate rises to 32% to 50%.³⁴

Reinfection or relapse?

Recurrence is one of the most frustrating and challenging complications of CDAD.

**Quinolones
may be more
strongly linked
to CDAD than
other
antibiotics**

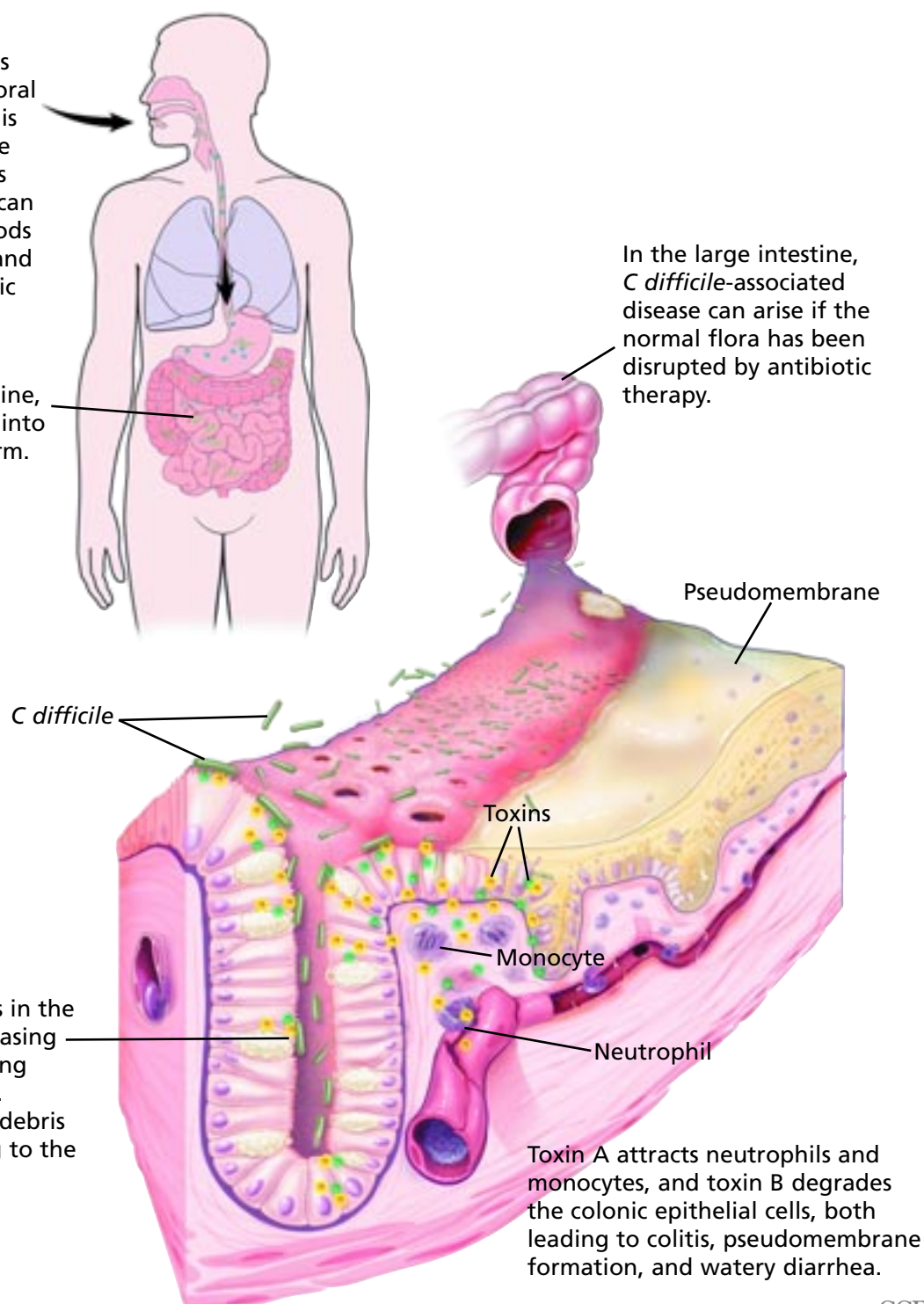
■ Pathogenesis of *C difficile*-associated disease

Clostridium difficile is spread via the fecal-oral route. The organism is ingested either as the vegetative form or as hardy spores, which can survive for long periods in the environment and can traverse the acidic stomach.

In the small intestine, spores germinate into the vegetative form.

In the large intestine, *C difficile*-associated disease can arise if the normal flora has been disrupted by antibiotic therapy.

C difficile reproduces in the intestinal crypts, releasing toxins A and B, causing severe inflammation. Mucous and cellular debris are expelled, leading to the formation of pseudomembranes.



Toxin A attracts neutrophils and monocytes, and toxin B degrades the colonic epithelial cells, both leading to colitis, pseudomembrane formation, and watery diarrhea.

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Medical Illustrator: David Schumick ©2006

FIGURE 3

TABLE 1

Advantages and disadvantages of diagnostic testing methods for *C difficile*

DIAGNOSTIC TEST	TURN-AROUND TIME	SENSITIVITY	ADVANTAGES	DISADVANTAGES
Endoscopy	2 hours	51%	Diagnostic of pseudomembranous colitis	Low sensitivity
Anaerobic culture	72 hours	89%–100%	Results useful for molecular typing	Does not distinguish toxin-producing strains
Tissue cytotoxic assay	48 hours	94%–100%	Detects A-B+ strains Gold standard	False-positives Results vary with experience of the technologist
Common antigen	15–45 minutes	58%–92%	Detects A-B+ strains Easy to use	Does not distinguish toxin-producing strains Cross-reacts with other anaerobes
Enzyme-linked immunosorbent assay (ELISA)—toxin A	2 hours	80%–95%	Easy to use	Does not detect A-B+ strains
ELISA—toxin A + B	2 hours		Detects A-B+ strains	Increased sensitivity for low-level toxin production
Immunochromatographic toxin A	< 1 hour	60%–85%	Simple to use Rapid	Does not detect A-B+ strains

BASED ON DATA FROM KELLY CP, POTHOUKAKIS C, LAMONT JT. *CLOSTRIDIUM DIFFICILE* COLITIS. N ENGL J MED 1994; 330:257–262; GERDING DN, JOHNSON S, PETERSON LR, MULLIGAN ME, SILVA J, JR. *CLOSTRIDIUM DIFFICILE*-ASSOCIATED DIARRHEA AND COLITIS. INFECT CONTROL HOSP EPIDEMIOL 1995; 16:459–477; AND WILKINS TD, LYERLY DM. *CLOSTRIDIUM DIFFICILE* TESTING: AFTER 20 YEARS, STILL CHALLENGING. J CLIN MICROBIOL 2003; 41:531–534

There is no universal agreement on how to clinically distinguish whether a second episode of CDAD is a reinfection or a relapse. One definition of a relapse is a recurrence of symptoms within 2 months of CDAD diagnosis; a reinfection is a recurrence of symptoms after 2 months.^{35–37} However, studies of patients who were thought to have had a relapse within 2 months of a previous CDAD episode indicate that 48% to 56% were actually reinfected with a different strain of *C difficile*.^{36,37}

Be it a reinfection or a true relapse, 12% to 24% of patients develop a second episode of CDAD within 2 months of the initial diagnosis. If a patient has two or more episodes of CDAD, the risk of additional recurrences

increases to 50% to 65%.³⁴

These statistics highlight the importance of preventive strategies (see below).

■ DIAGNOSIS

C difficile should be suspected in any adult with antimicrobial-associated diarrhea, and CDAD can occur up to several months after antimicrobial treatment is ended.^{26,33}

Only watery or loose stools should be tested for *C difficile* because the rate of colonization is high: a positive result in a normal stool sample proves that the patient is colonized with *C difficile* but not necessarily infected.³⁸ The primary exception to this rule is when you suspect CDAD in a patient with intestinal

ileus, which occurs in fewer than 1% of cases. Since most laboratories will not accept solid stool for *C difficile* testing, the clinician should notify the laboratory of the specific circumstances of the patient.

In general, empiric therapy without testing for *C difficile* is inappropriate, since only 30% of hospitalized patients with diarrhea have CDAD, even in an epidemic setting. Exceptions include severely ill or rapidly deteriorating patients at high risk for CDAD, in whom empiric therapy may be appropriate while awaiting test results.

There are a variety of tests for *C difficile*, each with advantages and disadvantages (TABLE 1). Factors to consider when selecting a diagnostic test include turnaround time, sensitivity, specificity, cost, whether there is an ongoing outbreak, and, of course, availability.

Enzyme immunoassays, available in most clinical laboratories, are fast and require less technical expertise than tissue culture. Although the negative predictive value hinges on the sensitivity of the particular assay, in most cases one negative result is enough to rule out CDAD. Nonetheless, a high clinical suspicion may warrant repeat testing.

Anaerobic bacterial culture is the method employed least by hospitals to diagnose CDAD, owing to its cost and turnaround time of approximately 72 hours.³⁹ In addition, this method's accuracy varies considerably in different laboratories because the methods and culture media are not standardized. The primary advantage of anaerobic culture is that it lends itself to molecular typing of strains, which may be useful in an outbreak.

■ TREATMENT

Stop the inciting antibiotic

Stopping the inciting antibiotic is the most important step in the initial treatment of CDAD.³⁶ Up to 25% of patients with CDAD recover without further therapy; in a series from 1974,⁴⁰ before there was effective therapy for CDAD, all 20 patients with pseudomembranous colitis eventually recovered after clindamycin treatment was stopped.

Oral metronidazole for mild disease; vancomycin for severe

In addition to stopping the inciting antibiotic, appropriate oral antimicrobial therapy directed specifically against *C difficile* should be given for 10 days to treat mild to moderate CDAD.^{41,42} In a study of 189 patients with CDAD, 97% responded to initial antibiotic therapy.⁴³

Although most patients in this study received oral vancomycin, several older studies comparing oral metronidazole to oral vancomycin for the treatment of CDAD indicate that metronidazole has been, at least historically, as effective as oral vancomycin and less expensive.^{44,45} In addition, widespread use of oral vancomycin could lead to vancomycin resistance. For these reasons, most experts recommend metronidazole as the first-line antimicrobial therapy for CDAD.^{41,46}

Recently, however, a prospective observational study⁴⁷ reported that the response rate with metronidazole was only 78%—significantly lower than previously published rates of response to oral vancomycin and oral metronidazole.

In an accompanying editorial to that report, Dr. Dale Gerding⁴² commented on the significance of these and other data, taking into account the recent emergence of a more virulent strain of *C difficile* as described above.²⁹ He concluded that metronidazole is an appropriate first-line treatment for most cases of CDAD, provided that the clinician is vigilant about monitoring the response to therapy. He indicated, however, that an alternative first-line therapy, such as oral or intraluminal vancomycin, should be considered for patients who present with moderate or severe disease.

CDAD can progress quickly

Along the same lines, it is important to realize that mild CDAD can quickly progress to moderate or severe disease and that these distinctions are not always easy to make.

Specific signs and symptoms of moderate disease may include fever, profuse diarrhea, abdominal pain, and leukocytosis.⁴¹ Severe disease is defined as the presence of complications of colitis, such as sepsis, volume depletion, electrolyte imbalance, hypotension, peritonitis, paralytic ileus, and toxic

No regimen has proven more effective than oral vancomycin or metronidazole

TABLE 2

Therapeutic options for *C difficile*-associated disease

DISEASE/HOST CHARACTERISTICS	RECOMMENDED THERAPY
Mild disease (No systemic symptoms, only mild diarrhea)	Metronidazole 250 mg by mouth four times a day or 500 mg by mouth three times a day for 10 days
Moderate disease (Fever, profuse diarrhea, abdominal pain, leukocytosis)	Vancomycin 125–500 mg by mouth four times a day for 10 days
Severe disease (Paralytic ileus, toxic megacolon, dehydration or sepsis)	Surgical consult plus intraluminal vancomycin
Inability to take oral medications	Intraluminal vancomycin with or without intravenous metronidazole

BASED ON DATA FROM MALNICK SD, ZIMHONY O. TREATMENT OF *CLOSTRIDIUM DIFFICILE*-ASSOCIATED DIARRHEA. ANN PHARMACOTHER 2002; 36:1767–1775 AND APISARNTHANARAK A, KHOURY H, REINUS WR, CRIPPIN JS, MUNDY LM. SEVERE *CLOSTRIDIUM DIFFICILE* COLITIS: THE ROLE OF INTRACOLONIC VANCOMYCIN? AM J MED 2002; 112:328–329.

megacolon.⁴¹ Some also include a white blood cell count of greater than $20 \times 10^9/L$ and elevated creatinine as indicators of severe disease.^{42,48}

Patients with signs of severe disease should receive oral vancomycin as initial therapy.

Consider surgery if CDAD progresses

Because CDAD can progress despite appropriate therapy, the clinician should follow the patient closely to see if symptoms improve within 1 to 2 days of starting therapy.⁴² Fever should subside within 24 to 48 hours and diarrhea should resolve within 2 to 5 days.^{42,43} If the disease progresses after starting treatment, additional or alternative therapeutic options should be considered, including a surgical consult for any signs of toxic megacolon, peritonitis, or sepsis (TABLE 2). However, if the patient's condition does not deteriorate, one should not conclude that treatment has failed before 6 to 7 days of therapy.⁴²

If oral therapy cannot be given

In some circumstances, oral therapy cannot be given, especially in severely ill or postoperative patients. In these situations, intracolonic vancomycin has been shown to be effective.⁴⁹ The role of intravenous metronidazole for

CDAD has yet to be determined, although high concentrations of metronidazole have been found in the stool after intravenous administration.⁵⁰

Alternative regimens

Many alternative regimens for CDAD have been explored, including different dosing strategies for vancomycin, other antimicrobials, probiotics, bacteriotherapy, adsorbents, and immunotherapy.³⁴ No regimen has proven to be significantly more effective than oral vancomycin or metronidazole for first-line therapy.

Some studies suggest, however, that treatment of recurrent CDAD with pulsed or tapered dosing of oral vancomycin may reduce recurrence rates.^{34,51} Some evidence also supports the use of probiotics such as *Saccharomyces boulardii* or *Lactobacillus* species in conjunction with vancomycin or metronidazole to reduce the recurrence rate of CDAD.³⁴

In 84 reported cases,³⁴ fecal enemas were given to replace the microflora disrupted by *C difficile* and antimicrobials; the recurrence rate was 10%. However, there have been no randomized controlled trials of this strategy.

Adsorbents such as ion-exchange resins and polymers, which, in theory, bind the *C*

Vigilant hand-washing and isolation precautions are key to controlling *C difficile*

difficile toxins in the colonic lumen before they can attach to enterocytes and cause disease, have also been tried. Although studies in animals were promising, human studies have not shown currently available agents to be superior to standard therapies. In fact, cholestyramine has actually been shown to bind to vancomycin, leading to suboptimal drug levels.

Intravenous immunoglobulin has demonstrated some positive results according to case reports, but no randomized controlled trials have been done.³⁴

Avoid antiperistaltic agents

Antiperistaltic agents should not be given, either alone or in conjunction with other therapy.^{5,6} This recommendation is based on anecdotal data indicating that diphenoxylate and atropine may predispose patients with CDAD to toxic megacolon. Since narcotics also have antiperistaltic effects, they too should be avoided in patients with CDAD.^{5,6} Narcotics may have contributed to the poor outcome of the case described at the beginning of this article.

Gauge response to therapy on signs and symptoms

Therapeutic response should be based purely on clinical signs and symptoms: a repeat toxin assay should not be done as a “test of cure,” since patients may remain colonized with toxin-producing strains following recovery.⁵²

Do not treat asymptomatic colonization

Current therapies are ineffective for eradicating asymptomatic colonization. Vancomycin has been studied for this purpose. However, experts do not recommend treating patients colonized with *C difficile* as an infection control strategy.⁴² Its effects are not sustained and patients may be at increased risk for prolonged carriage after treatment ends.

Significant intraluminal levels of metronidazole are achieved only in the presence of diarrhea, which renders the drug ineffective for patients with asymptomatic colonization.³⁶

■ PREVENTION

Two approaches to the prevention of CDAD include infection control, thus interrupting

the horizontal spread of *C difficile* within health care facilities, and reducing the individual patient's risk of acquiring the disease once exposed to the organism.³⁸ Probiotic agents have been studied as prophylaxis in patients receiving antimicrobial agents, but no statistically significant difference in rates of CDAD has been seen.³⁸

Contact precautions

Spread of *C difficile* in health care facilities has been well documented, occurring primarily person to person (from people with or without symptoms) and via contamination of the patient care environment.^{38,53} The most effective means of decreasing horizontal spread of *C difficile* has been a combination of vigilant hand hygiene and use of isolation precautions.³⁶

The literature contains both direct and indirect evidence for contamination of health care workers' hands in endemic and outbreak settings. Alcohol is not effective in killing *C difficile* spores. Therefore, if a hospital is experiencing an outbreak, it is prudent for health care workers to wash their hands exclusively with soap and water when caring for patients with known CDAD.⁵⁴

The 1994 Hospital Infection Control Practices Advisory Committee (HICPAC) Guideline for Isolation Precautions in Hospitals recommends contact precautions for symptomatic patients. These include placing patients in private rooms or cohorting (grouping patients in a designated area) and donning gowns and gloves when entering the patient's room.⁵⁵ One hospital reported a 60% decrease in CDAD incidence after instituting a more stringent infection control program, including increased enforcement of contact precautions, a monthly educational program, triclosan-containing hand soap, and increased environmental cleaning.⁵⁶

Bleach for environmental disinfection

Environmental contamination of *C difficile* is due to persistence of spores that can be highly resistant to routine disinfectants and can survive on dry surfaces for many weeks or months. The rate of surface contamination increases in proportion to the *C difficile* status, severity of diarrhea, and incontinence of

patients in the area. Environments of asymptomatic carriers have lower rates than those of patients with symptomatic disease.³⁸

Patient-care items such as reusable electronic rectal thermometers have been implicated in outbreaks, and dedication of single-use items to individual patients can eliminate this source of contamination.^{24,38} "High-touch" surfaces in patients' bathrooms (eg, light switches) have also been implicated in outbreaks and should be targeted for enhanced environmental cleaning.

No well-controlled trials of disinfectants have been conducted; however, use of both unbuffered and phosphate-buffered hypochlorite solutions (bleach) has been shown to decrease rates of *C difficile* contamination, and some studies suggest that cleaning with bleach may lower CDAD rates.^{57–59} Although no disinfectants are registered with the Environmental Protection Agency with a claim for *C difficile* spore inactivation, the HICPAC Guideline for Environmental Infection Control in Healthcare Facilities recommends "meticulous cleaning followed by disinfection

using hypochlorite-based germicides as appropriate."⁶⁰ Dilutions and schedule of mixing bleach solutions for this purpose can be found in the HICPAC guideline.

Restrict antibiotic use?

Since prior antimicrobial use is associated with the vast majority of patients who develop health-care-associated CDAD, restricting the use of specific antimicrobial agents would seem to be an important infection-control strategy to reduce patient risk. Unfortunately, with the exception of clindamycin restriction, few reports demonstrate success of this approach.

However, reduction of unnecessary antimicrobial use in general would reduce the risk of CDAD in all patients. Clinicians who have treated patients with severe or recurrent CDAD may gain an increased appreciation of the serious risks associated with unnecessary antimicrobial therapy. ■

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REFERENCES

1. Bignardi GE. Risk factors for *Clostridium difficile* infection. *J Hosp Infect* 1998; 40:1–15.
2. Dial S, Alrasadi K, Manoukian C, Huang A, Menzies D. Risk of *Clostridium difficile* diarrhea among hospital inpatients prescribed proton pump inhibitors: cohort and case-control studies. *CMAJ* 2004; 171:33–38.
3. Pepin J, Saheb N, Coulombe M-A, et al. Emergence of fluoroquinolones as the predominant risk factor for *Clostridium difficile*-associated diarrhea: a cohort study during an epidemic in Quebec. *Clin Infect Dis* 2005; 41:1254–1260.
4. Dial S, Delaney JAC, Barkun AN, Suissa S. Use of gastric acid-suppressive agents and the risk of community-acquired *Clostridium difficile*-associated disease. *JAMA* 2005; 294:2989–2995.
5. George WL, Rolfe RD, Finegold SM. Treatment and prevention of antimicrobial agent-induced colitis and diarrhea. *Gastroenterology* 1980; 79:366–372.
6. Cone JB, Wetzel W. Toxic megacolon secondary to pseudomembranous colitis. *Dis Colon Rectum* 1982; 25:478–482.
7. Larson HE, Price AB, Honour P, Borriello SP. *Clostridium difficile* and the aetiology of pseudomembranous colitis. *Lancet* 1978; 1:1063–1066.
8. Bartlett JG, Chang TW, Gurwith M, Gorbach SL, Onderdonk AB. Antibiotic-associated pseudomembranous colitis due to toxin-producing clostridia. *N Engl J Med* 1978; 298:531–534.
9. Lyerly DM, Krivan HC, Wilkins TD. *Clostridium difficile*: its disease and toxins. *Clin Microbiol Rev* 1988; 1:1–18.
10. Gerding DN. Disease associated with *Clostridium difficile* infection. *Ann Intern Med* 1989; 110:255–257.
11. McFarland LV, Stamm WE. Review of *Clostridium difficile*-associated diseases. *Am J Infect Control* 1986; 14:99–109.
12. Bartlett JG. *Clostridium difficile*: history of its role as an enteric pathogen and the current state of knowledge about the organism. *Clin Infect Dis* 1994; 18(suppl 4):S265–S272.
13. Archibald LK, Banerjee SN, Jarvis WR. Secular trends in hospital-acquired *Clostridium difficile* disease in the United States, 1987–2001. *J Infect Dis* 2004; 189:1585–1589.
14. McDonald LC, Banerjee S, Jernigan DB. Increasing incidence of *Clostridium difficile*-associated disease in U.S. acute care hospitals, 1993–2001 [Abstract 45]. 04 Apr 18; Alexandria, VA: Society for Healthcare Epidemiology of America; 2004.
15. Olson MM, Shanholtzer CJ, Lee JT Jr, Gerding DN. Ten years of prospective *Clostridium difficile*-associated disease surveillance and treatment at the Minneapolis VA Medical Center, 1982–1991. *Infect Control Hosp Epidemiol* 1994; 15:371–381.
16. Miller MA, Hyland M, Ofner-Agostini M, Gourdeau M, Ishak M. Morbidity, mortality, and healthcare burden of nosocomial *Clostridium difficile*-associated diarrhea in Canadian hospitals. *Infect Control Hosp Epidemiol* 2002; 23:137–140.
17. Kyne L, Hamel MB, Polavaram R, Kelly CP. Health care costs and mortality associated with nosocomial diarrhea due to *Clostridium difficile*. *Clin Infect Dis* 2002; 34:346–353.
18. Wilcox MH, Cuniffe JG, Trundle C, Redpath C. Financial burden of hospital-acquired *Clostridium difficile* infection. *J Hosp Infect* 1996; 34:23–30.
19. Pepin J, Valiquette L, Cossette B. Mortality attributable to nosocomial *Clostridium difficile*-associated disease during an epidemic caused by a hypervirulent strain in Quebec. *Can Med J* 2005; 173. Online advance.
20. Barbut F, Petit JC. Epidemiology of *Clostridium difficile*-associated infections. *Clin Microbiol Infect* 2001; 7:405–410.
21. Gaynes R, Rimland D, Killum E, et al. Outbreak of *Clostridium difficile* infection in a long-term care facility: association with gatifloxacin use. *Clin Infect Dis* 2004; 38:640–645.
22. Loo VG, Poirier L, Miller MA, et al. A predominantly clonal multi-institutional outbreak of *Clostridium difficile*-associated diarrhea with high morbidity and mortality. *N Engl J Med* 2005;

353:2442–2449.

23. Kelly CP, Pothoulakis C, Lamont JT. *Clostridium difficile* colitis. N Engl J Med 1994; 330:257–262.
24. Brooks SE, Veal RO, Kramer M, Dore L, Schupf N, Adachi M. Reduction in the incidence of *Clostridium difficile*-associated diarrhea in an acute care hospital and a skilled nursing facility following replacement of electronic thermometers with single-use disposables. Infect Control Hosp Epidemiol 1992; 13:98–103.
25. McNulty C, Logan M, Donald IP, et al. Successful control of *Clostridium difficile* infection in an elderly care unit through use of a restrictive antibiotic policy. J Antimicrob Chemother 1997; 40:707–711.
26. Simor AE, Bradley SF, Strausbaugh LJ, Crossley K, Nicolle LE. *Clostridium difficile* infection in long-term-care facilities for the elderly. Infect Control Hosp Epidemiol 2002; 23:696–703.
27. Simor AE, Yake SL, Tsimidis K. Infection due to *Clostridium difficile* among elderly residents of a long-term-care facility. Clin Infect Dis 1993; 17:672–678.
28. Eglow R, Pothoulakis C, Itzkowitz S, et al. Diminished *Clostridium difficile* toxin A sensitivity in newborn rabbit ileum is associated with decreased toxin A receptor. J Clin Invest 1992; 90:822–829.
29. Warny M, Pepin J, Fang A, et al. Toxin production by an emerging strain of *Clostridium difficile* associated with outbreaks of severe disease in North America and Europe. Lancet 2005; 366:1079–1084.
30. McDonald LC, Killgore GE, Thompson A, et al. An epidemic, toxin gene-variant strain of *Clostridium difficile*. N Engl J Med 2005; 353:2433–2441.
31. Shim JK, Johnson S, Samore MH, Bliss DZ, Gerding DN. Primary symptomless colonisation by *Clostridium difficile* and decreased risk of subsequent diarrhoea. Lancet 1998; 351:633–636.
32. Kyne L, Warny M, Qamar A, Kelly CP. Asymptomatic carriage of *Clostridium difficile* and serum levels of IgG antibody against toxin A. N Engl J Med 2000; 342:390–397.
33. Palmore TN, Sohn S, Malak SF, Eagan J, Sepkowitz KA. Risk factors for acquisition of *Clostridium difficile*-associated diarrhea among outpatients at a cancer hospital. Infect Control Hosp Epidemiol 2005; 26:680–684.
34. McFarland LV. Alternative treatments for *Clostridium difficile* disease: what really works? J Med Microbiol 2005; 54:101–111.
35. Pepin J, Alary ME, Valiquette L, et al. Increasing risk of relapse after treatment of *Clostridium difficile* colitis in Quebec, Canada. Clin Infect Dis 2005; 40:1591–1597.
36. Barbut F, Richard A, Hamadi K, Chomette V, Burghoffer B, Petit JC. Epidemiology of recurrences or reinfections of *Clostridium difficile*-associated diarrhea. J Clin Microbiol 2000; 38:2386–2388.
37. Wilcox MH, Fawley WN, Settle CD, Davidson A. Recurrence of symptoms in *Clostridium difficile* infection—relapse or reinfection? J Hosp Infect 1998; 38:93–100.
38. Gerding DN, Johnson S, Peterson LR, Mulligan ME, Silva J Jr. *Clostridium difficile*-associated diarrhea and colitis. Infect Control Hosp Epidemiol 1995; 16:459–477.
39. Wilkins TD, Lyerly DM. *Clostridium difficile* testing: after 20 years, still challenging. J Clin Microbiol 2003; 41:531–534.
40. Tedesco FJ, Barton RW, Alpers DH. Clindamycin-associated colitis. A prospective study. Ann Intern Med 1974; 81:429–433.
41. Malnick SD, Zimhony O. Treatment of *Clostridium difficile*-associated diarrhea. Ann Pharmacother 2002; 36:1767–1775.
42. Gerding DN. Metronidazole for *Clostridium difficile*-associated disease: is it okay for Mom? Clin Infect Dis 2005; 40:1598–1600.
43. Bartlett JG. Treatment of antibiotic-associated pseudomembranous colitis. Rev Infect Dis 1984; 6(suppl 1):S235–S241.
44. Teasley DG, Gerding DN, Olson MM, et al. Prospective randomised trial of metronidazole versus vancomycin for *Clostridium difficile*-associated diarrhoea and colitis. Lancet 1983; 2:1043–1046.
45. Wenisch C, Parschalk B, Hasenhundl M, Hirschl AM, Graninger W. Comparison of vancomycin, teicoplanin, metronidazole, and fusidic acid for the treatment of *Clostridium difficile*-associated diarrhea. Clin Infect Dis 1996; 22:813–818.
46. Centers for Disease Control and Prevention. Recommendations for preventing the spread of vancomycin resistance: recommendations of the Hospital Infection Control Practices Advisory Committee (HIC-PAC). MMWR 1995; 44(No. RR-12):1–13.
47. Musher DM, Aslam S, Logan N, et al. Relatively poor outcome after treatment of *Clostridium difficile* colitis with metronidazole. Clin Infect Dis 2005; 40:1586–1590.
48. Pepin J, Valiquette L, Alary ME, et al. *Clostridium difficile*-associated diarrhea in a region of Quebec from 1991 to 2003: a changing pattern of disease severity. CMAJ 2004; 171:466–472.
49. Apisarnthanarak A, Khoury H, Reinus WR, Crippin JS, Mundy LM. Severe *Clostridium difficile* colitis: the role of intracolonic vancomycin? Am J Med 2002; 112:328–329.
50. Bolton RP, Culshaw MA. Faecal metronidazole concentrations during oral and intravenous therapy for antibiotic associated colitis due to *Clostridium difficile*. Gut 1986; 27:1169–1172.
51. McFarland LV, Elmer GW, Surawicz CM. Breaking the cycle: treatment strategies for 163 cases of recurrent *Clostridium difficile* disease. Am J Gastroenterol 2002; 97:1769–1775.
52. Poutanen SM, Simor AE. *Clostridium difficile*-associated diarrhea in adults. CMAJ 2004; 171:51–58.
53. McFarland LV, Mulligan ME, Kwok RY, Stamm WE. Nosocomial acquisition of *Clostridium difficile* infection. N Engl J Med 1989; 320:204–210.
54. Boyce JM, Pittet D. Guideline for hand hygiene in health-care settings: recommendations of the Healthcare Infection Control Practices Advisory Committee and the HICPAC/SHEA/APIC/IDSA Hand Hygiene Task Force. Infect Control Hosp Epidemiol 2002; 23(12 suppl):S3–40.
55. Garner JS. Guideline for isolation precautions in hospitals. The Hospital Infection Control Practices Advisory Committee. Infect Control Hosp Epidemiol 1996; 17:53–80.
56. Zafar AB, Gaydos LA, Furlong WB, Nguyen MH, Mennonna PA. Effectiveness of infection control program in controlling nosocomial *Clostridium difficile*. Am J Infect Control 1998; 26:588–593.
57. Kaatz GW, Gitlin SD, Schaberg DR, et al. Acquisition of *Clostridium difficile* from the hospital environment. Am J Epidemiol 1988; 127:1289–1294.
58. Mayfield JL, Leet T, Miller J, Mundy LM. Environmental control to reduce transmission of *Clostridium difficile*. Clin Infect Dis 2000; 31:995–1000.
59. Wilcox MH, Fawley WN, Wigglesworth N, Parnell P, Verity P, Freeman J. Comparison of the effect of detergent versus hypochlorite cleaning on environmental contamination and incidence of *Clostridium difficile* infection. J Hosp Infect 2003; 54:109–114.
60. Sehulster L, Chinn RY. Guidelines for environmental infection control in health-care facilities. Recommendations of CDC and the Healthcare Infection Control Practices Advisory Committee (HICPAC). MMWR Recomm Rep 2003; 52(RR-10):1–42.

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